

BACKGROUND

Fatty liver disease (FLD), often caused by high-fat diets, is a major health concern. This study investigates:

- The role of fatty acid oxidative metabolism in FLD
- Potential of other fatty acid metabolism pathways as therapeutic targets for FLD

HYPOTHESIS

We hypothesize that mitochondrial fatty acid oxidation is crucial in FLD pathogenesis. Knocking out the CACT gene in mice will result in more severe FLD progression on a high-fat diet compared to controls, with consistent molecular markers of mitochondrial and peroxisomal dysfunction.

MATERIALS & METHODS

Gene Knockout

- CACT knockout mice were compared to wild-type C57BL/6J controls.

Fatty Liver Diet Regimen

Mice were divided into four groups:

- Control genotype / control diet (Control) (n=5)
- Control genotype / fatty liver diet (Control FLD) (n=4)
- CACT knockout / control diet (CactLiv^{-/-}) (n=4)
- CACT knockout / fatty liver diet (CactLiv^{-/-} FLD) (n=5)

Control diet: standard chow (10 kcal% fat)

Fatty liver diet: high-fat (60kcal% fat) for 7 days

Liver weight measured post-diet

RT-PCR Analysis

- Quantitative real-time PCR with SYBR Green on QuantStudio 6 Flex
- Primers for mitochondrial biosynthesis, gluconeogenesis, fatty acid oxidation, ketogenesis, triacylglycerol synthesis
- B2M as housekeeping gene
- Samples run in duplicate with melt curve analysis for specificity

Data Analysis

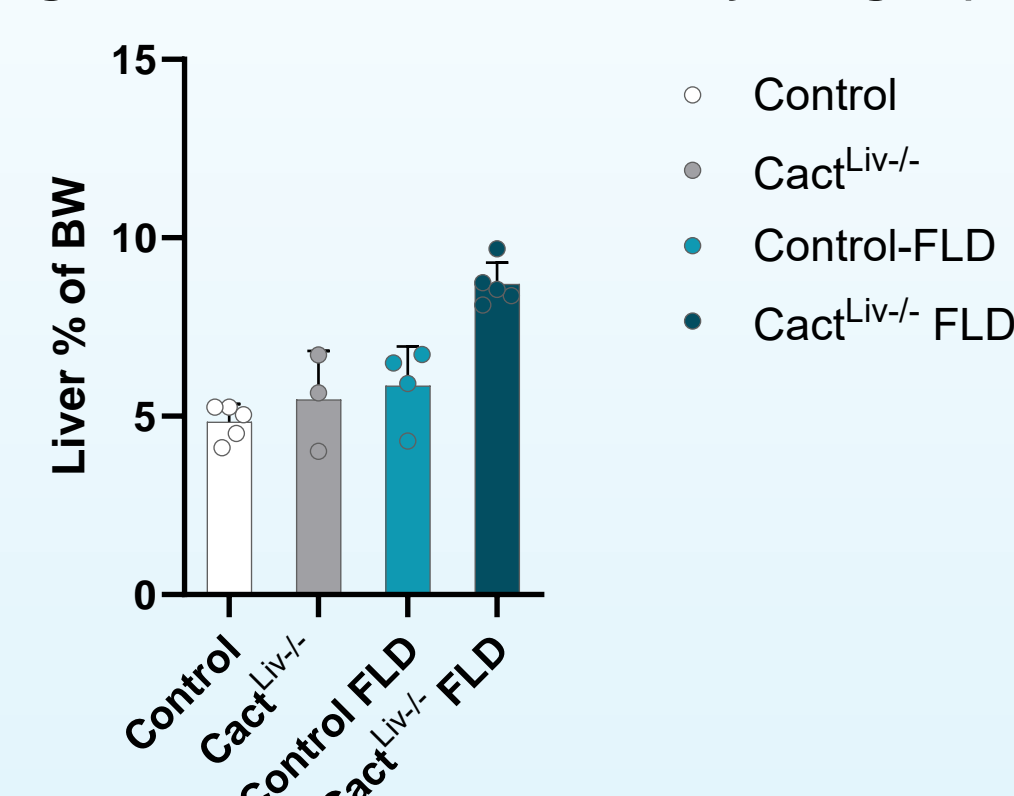
Relative gene expression: $2^{-\Delta\Delta Ct}$ method
Statistical analysis: GraphPad Prism, $p < 0.05$ considered significant

RESULTS

Liver weight:

- Increased in CactLiv^{-/-} FLD group

Fig. 1: Percentage Liver of Mouse Total Body Weight (BW)



Gene expression changes:

CactLiv^{-/-} mice:

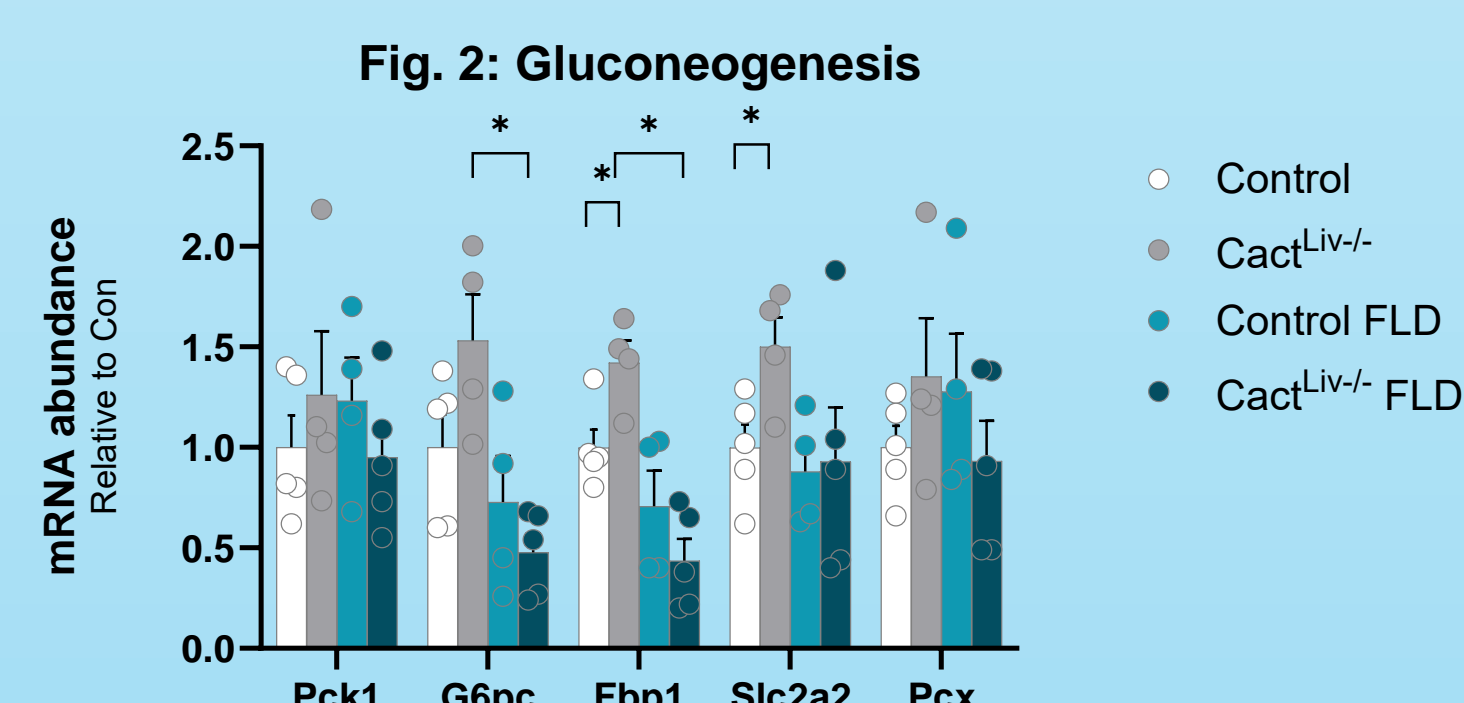
- ↑ Gluconeogenesis, mitochondrial/peroxisomal fatty acid oxidation, carnitine synthesis, ketogenesis

CactLiv^{-/-} FLD mice:

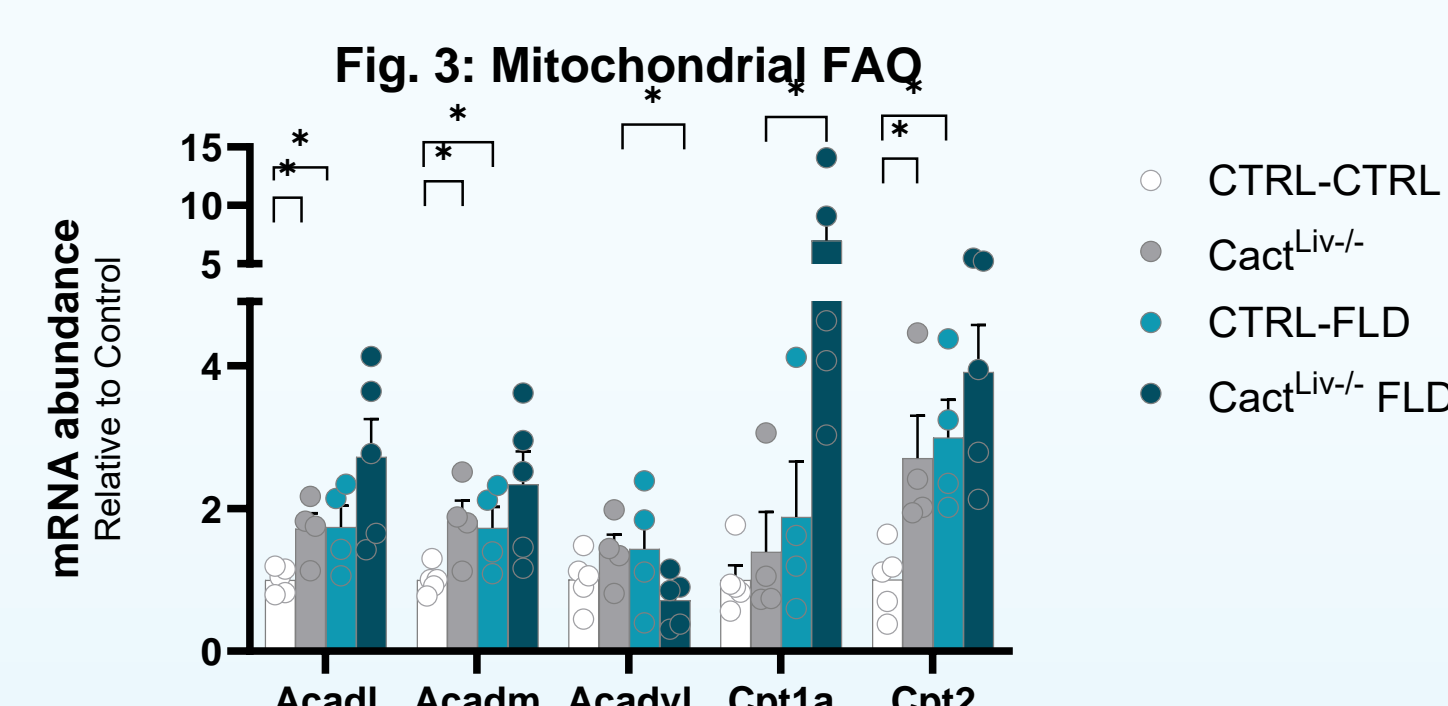
- ↑ Pparg (lipogenesis)
- ↓ Gluconeogenesis, carnitine synthesis, most mitochondrial fatty acid oxidation genes (except cpt1a)

Control FLD mice:

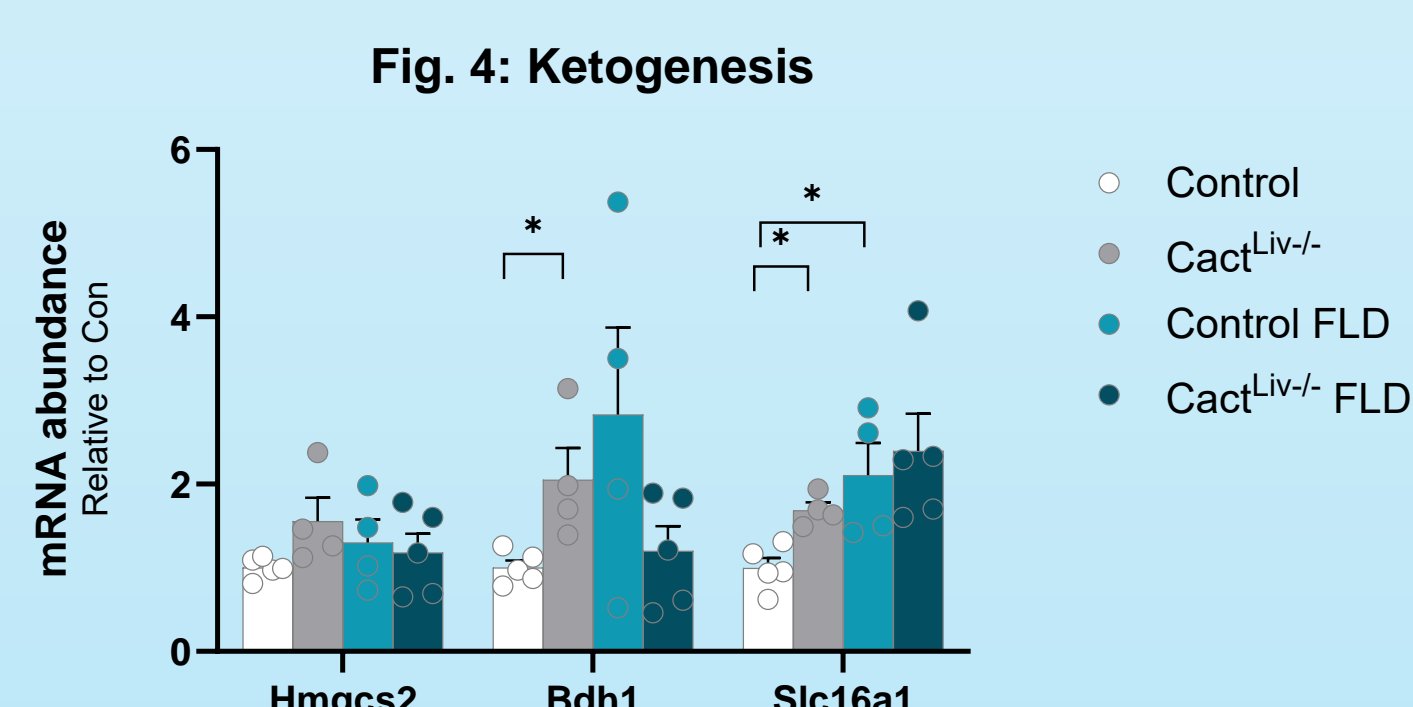
- ↑ Mitochondrial fatty acid oxidation, ketogenesis, systemic metabolism, peroxisomal fatty acid oxidation



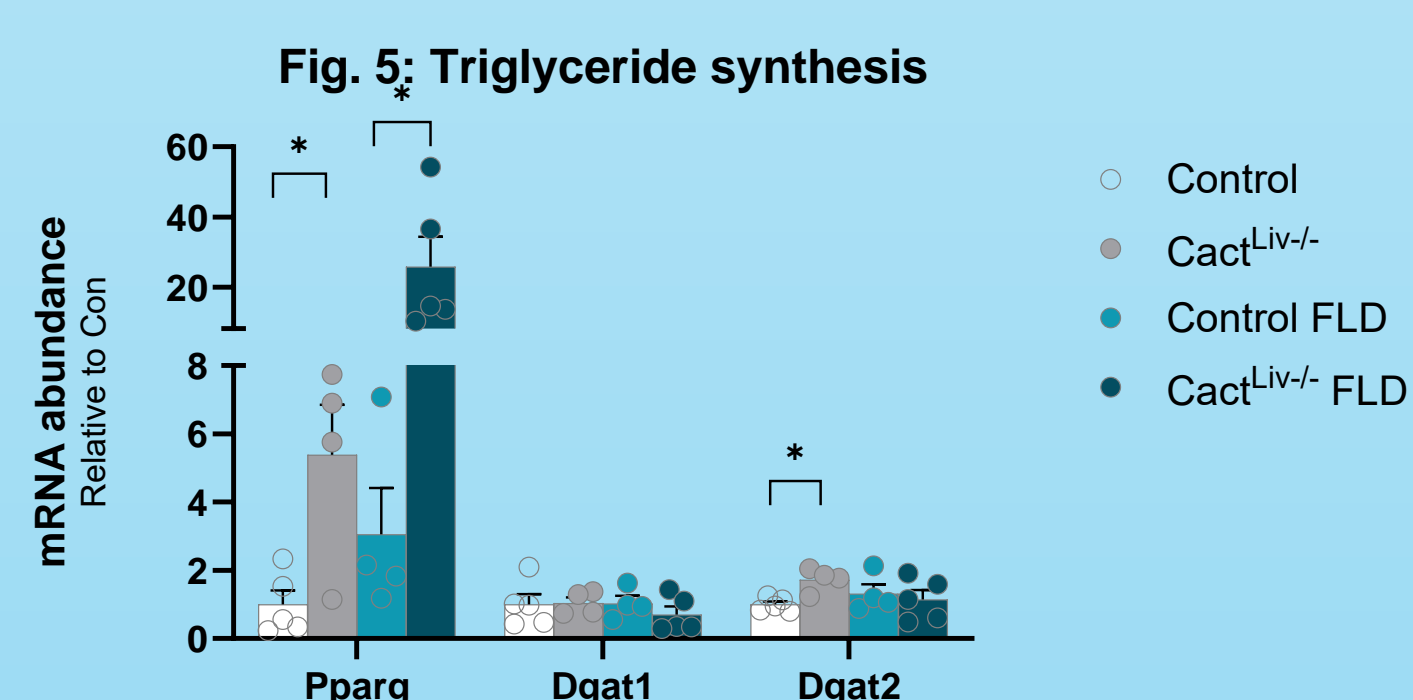
- Compared to Control, CactLiv^{-/-} mice show upregulation of pcx ($p=0.030$), and fbp1 ($p=0.021$). Yet, when compared to CactLiv^{-/-} mice, CactLiv^{-/-} FLD mice show downregulation of G6pc (p -value=0.002) and Fbp1 (p -value=0.0004).



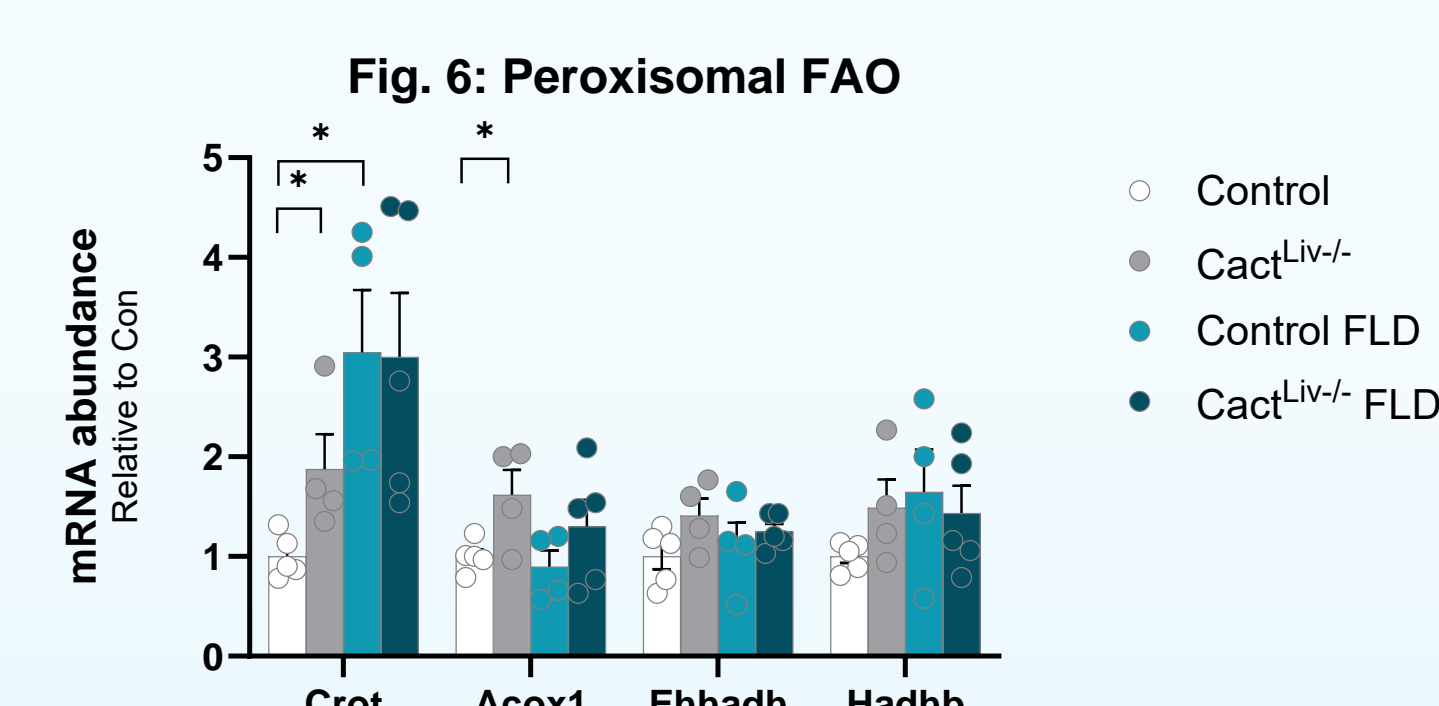
- Acadl ($p=0.012$), Cpt2 ($p=0.054$), and Acadm ($p=0.018$) present with increased abundance compared to control. Control shows moderate increase in expression of cpt2 (0.007) under FLD diet. CactLiv^{-/-} shows considerable upregulation of cpt1a ($p=0.051$) and moderate decrease in acadvl ($p=0.046$).



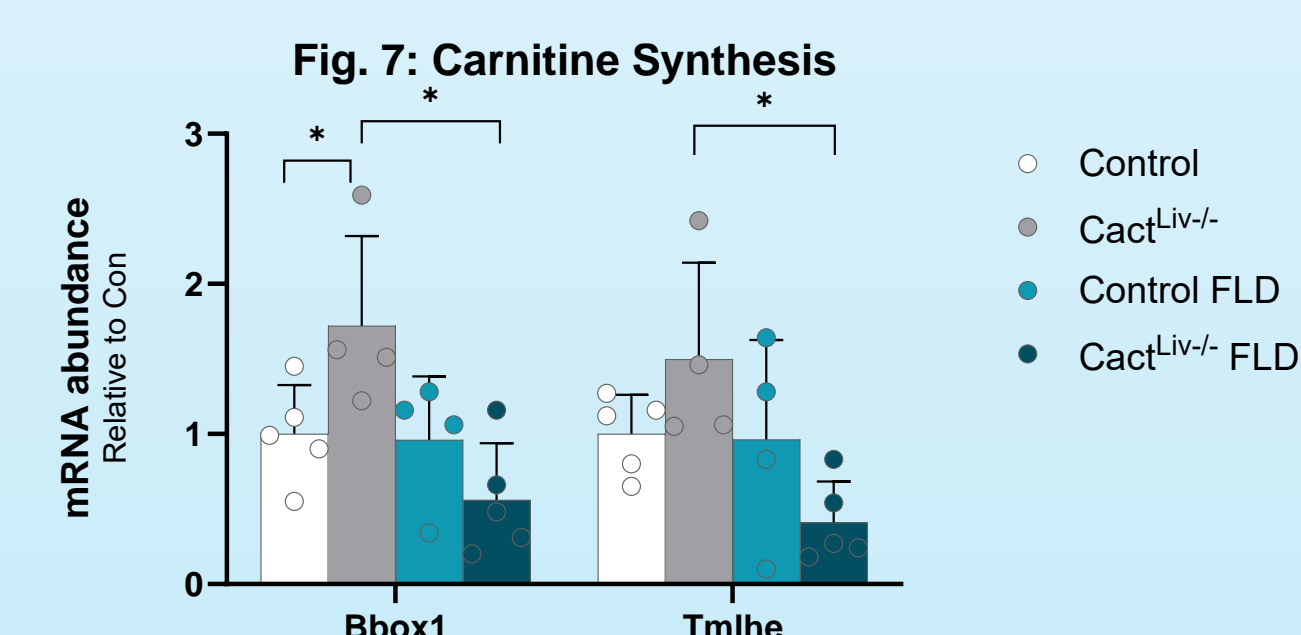
- CactLiv^{-/-} samples exhibit an increase of abundance for Slc16a1 ($p=0.003$), and bdh1 ($p=0.020$) when compared to Control. Slc16a1 also increased in Control FLD ($p=0.018$).



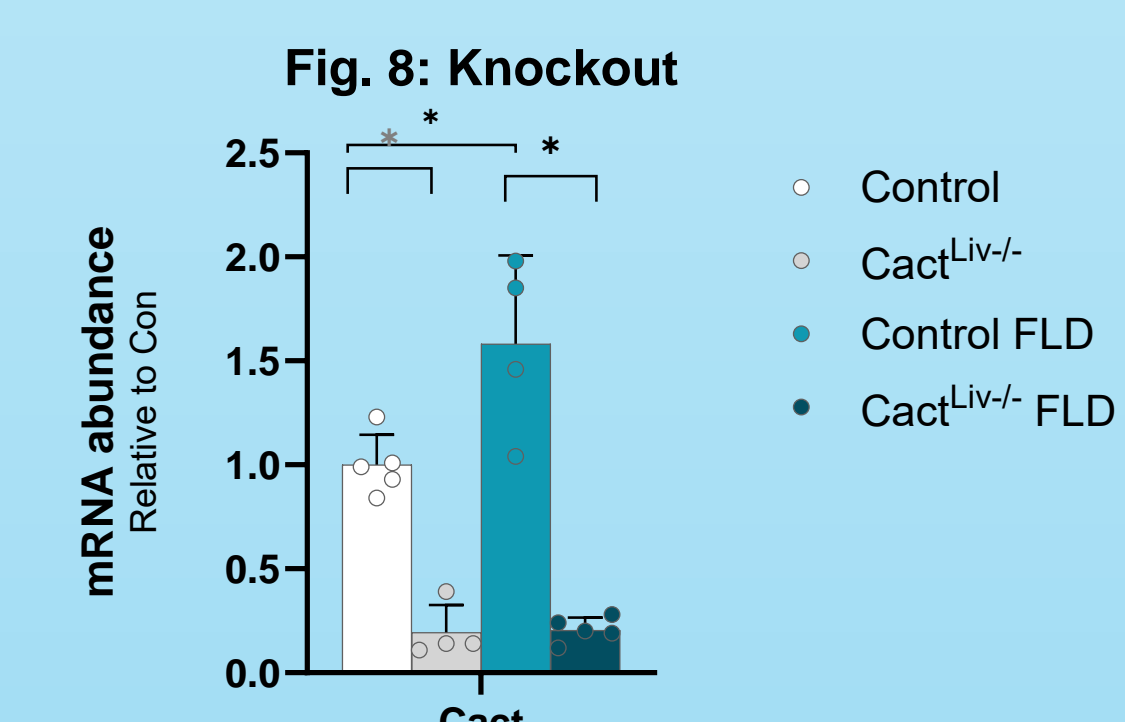
- Pparg ($p=0.015$) and Dgat2 ($p=0.006$) are seen with increased abundance in CactLiv^{-/-} mice. Pparg abundance is also seen to be greater in CactLiv^{-/-} FLD mice than Control FLD mice.



- CactLiv^{-/-} mice exhibit increased abundance of Crot ($p=0.032$) and acox1 ($p=0.034$) compared to Control. Crot is also seen in higher abundance in Control mice fed fatty liver inducing diet when compared to control diet ($p=0.008$).



- Bbox1 ($p=0.05$) is seen in higher abundance in CactLiv^{-/-} mice vs. Control. Yet, there is a significant decrease in abundance in Bbox1 ($p=0.009$) and Tmlhe ($p=0.011$) in CactLiv^{-/-} mice fed a fatty liver inducing diet, compared to CactLiv^{-/-} on a control diet.



- Gene Analysis shows that there is significant absence of Cact in knockout mice in Control vs CactLiv^{-/-} mice under control diet ($p=0.00006$) and FLD diet ($p=0.0002$). There is also a significant increase in Cact ($p=0.02$) in Control mice fed the FLD diet compared to those on control diet.

CONCLUSION/DISCUSSION

These findings suggest that mitochondrial fatty acid oxidation is crucial in protecting against FLD. CACT knockout impairs the liver's adaptive response to a high-fat diet, and alternative fatty acid oxidation pathways may be activated as compensatory mechanisms. Further research is needed to explore potential therapeutic targets based on these findings

Next Steps:

- Conduct a more comprehensive analysis of gene expression changes, possibly using RNA sequencing to identify additional pathways involved in the compensatory response to CACT knockout.
- Perform metabolomic studies to better understand the alterations in lipid metabolism and identify potential biomarkers.
- Investigate the effects of pharmacological interventions that target the upregulated pathways identified in this study, such as peroxisomal fatty acid oxidation, in CACT knockout mice.

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